

Epilithic Diatom Diversity in Golestan Waterfall

Jamileh Panahy Mirzahasanlou^{1*}, Tina Qarebesloum¹, Masoumeh Farasati², Arsalan Bahalkeh³

Received: 2020-05-20

Revised and accepted: 2020-09-15

Abstract

Waterfalls are considered as aquatic ecosystems, of which limited study was conducted on their biodiversity. Golestan waterfall is one of the outstanding waterfalls in eastern edge of Golestan National Park. Since no research was performed on algal flora in the area, the present study aimed to identify the epilithic diatoms of the waterfall, as a component of this ecosystem biodiversity. The samples were collected from stony substrates at each season. The results represented 24 genera including 47 species, which all taxa belonged to Bacillariophyceae. In addition, *Cymbopleura* with five species and *Gomphonema*, and *Navicula* with four species had more species, respectively. Further, *Achnantheidium minutissimum*, *Amphipleura pellucida*, *Cymbella affinis*, *Cymbopleura kuelbsii*, and *Gomphonema pumilum* were determined as the most abundant taxa. Furthermore, *Delicatophycus verena*, *Stauroneis separanda* and *Tryblionella brunoi* were found in the diatom flora of Iran for the first time. Due to the few diatom studies in Iran, conducting detailed and local

studies can improve data on diatom flora of Iran.

Keywords: Bacillariophyceae, *Cymbopleura*, Diatom Flora, Golestan Province, Iran

Introduction

Biodiversity plays a major role in ecosystem function and stability and is the life support system (Rawat and Agarwal, 2015). Diatoms are considered as an extremely diverse group of algae occurring in almost all aquatic systems. Additionally, they are a systematic group characterized by their siliceous wall. Taxonomic diversity in diatoms is important, which reflects the biodiversity and stability of an aquatic ecosystem (Andrejic et al., 2012). Further, their rapid response to environmental changes can represent the ecological conditions of the living environment, which makes them useful in water quality assessments (Delgado et al., 2012; Kelly et al., 2007; Martin et al., 2010; Noga et al., 2013).

Since 2016, some studies focused in Golestan province regarding diatoms in Zarringol

1-Department of Biology, Faculty of Basic Science, Gonbad Kavous University, Gonbad Kavous, Iran

2-Department of Range and Watershed Management, Faculty of Agriculture and Natural Resources, Gonbad Kavous University, Gonbad Kavous, Iran

3-Department of Fishery, Faculty of Agriculture and Natural Resources, Gonbad Kavous University, Gonbad Kavous, Iran

*E-mail address: panahi@gonbad.ac.ir

River (Dadgar, 2016), Chehal Chay River in Minudasht (Lakzaie et al., 2018), Gharah Chay River in Ramian (Bayani, 2019), three springs in Ramian (Ahmadi et al., 2019), and Khorrmarud River (Aghatabay et al., 2018). However, few studies have conducted on algal flora and diatom so far. Thus, the Further studies can enhance the knowledge on diatom flora in local and country scales. Waterfalls are interesting ecosystems, which are considered as vertical wetlands, and kept cool in summer and mild in winter. Many outstanding waterfalls exist in Golestan province, the algae of which have not been assessed floristically so far.

Material and methods

Golestan province with the area of 22033 km² is located in northeast Iran, southeast Caspian Sea, which has diverse climate and aquatic ecosystems due to its geographical position. Golestan waterfall is placed 47 km away from Galikesh city at the eastern

edge of Golestan National Park (Fig. 1). The present study sought to recognize the diatoms of the waterfall as a basic study. The diatoms were sampled from stony substrates by using toothbrush seasonally in three different sites and transferred to laboratory after fixation with 4% formalin (Bellinger and Sigeo, 2010). In addition, the samples were cleaned through acid digestion method (Taylor et al., 2007). Further, permanent slides were prepared using Canada Balsam and diatoms were identified by using a light microscope and considering available flora (Bahls, 2006, 2012, 2013; Krammer and Lange-Bertalot, 1986, 1988, 1991a, b; Krammer, 2002, 2003; Lange-Bertalot, 2001). Taxa names were checked in www.algaebase.org. In each slide, 300-400 valves were counted and the relative abundance of each taxon was calculated.

Along with algal samples, water ones were added into 1L containers and transferred to laboratory in order to analyze nitrate, am-

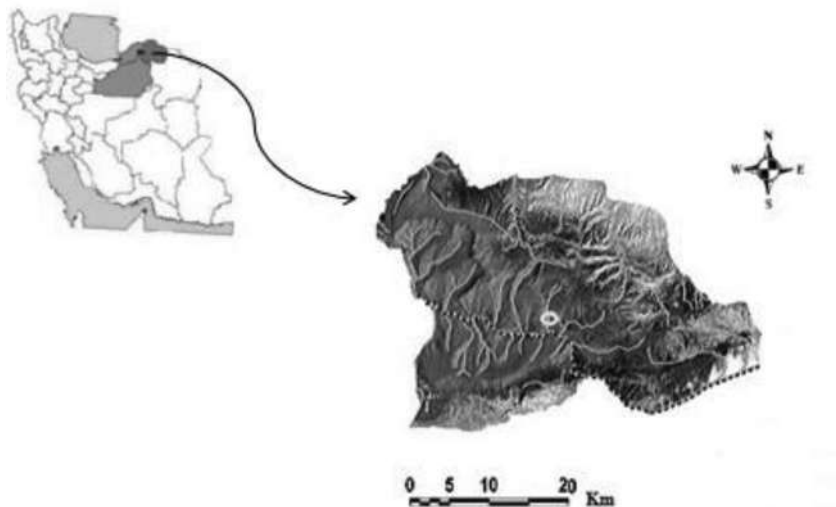


Fig. 1. Location of Golestan waterfall

monium, silicate, and phosphate (Clesceri et al., 1999). (Furthermore, dissolved oxygen (DO), temperature, EC, salinity, pH, and TDS were measured in situ by using a HQ40d portable device.

Results

Table 1 summarizes the physicochemical results related to water in different seasons. A total of 47 taxa belonging to 24 genera were identified in Golestan waterfall (Table 2, Plates I and II), all of which were related to the class Bacillariophyceae. Based on the results, the most species-rich genus included *Cymbopleura* with five species and *Gomphonema* and *Navicula* each with four species, followed by *Diploneis*, *Surirella*, and *Tryblionella* with three species (Fig. 2). Additionally, *A. minutissimum*, *Amphipleu-*

ra pellucida, *Cymbopleura kuelbsii*, *Gomphonema pumilum*, and *Cymbella affinis* were characterized as the most abundant taxa.

Based on the morphological groups, the identified taxa were classified into six groups, the highest number of which belonged to asymmetrical biraphid with 18 species and 8 genera, symmetrical biraphid with 13 species and 7 genera. Nitzschoid with 7 species and 3 genera, and Monoraphid with 3 species and 2 genera, respectively. Further, the least number was observed in Surirelloid with 3 species and 1 genera, and Araphids with 3 species in 3 genera, respectively.

In the present study, three species were determined as new for the diatom flora of Iran, which are provided as follows.

Table 1. Results related to water physicochemical parameters in Golestan waterfall during different seasons

Parameteres	Spring	Summer	Fall	Winter
PO ₄ (mg.l ⁻¹)	0.02	0.016	0.015	0.02
SiO ₂ (mg.l ⁻¹)	0.57	0.74	0.95	0.39
NO ₃ (mg.l ⁻¹)	0.61	2.33	1.42	0.55
NH ₃ (mg.l ⁻¹)	0.011	0.015	0.011	0.016
T (°C)	21.3	14.8	6.8	7
DO (mg.l ⁻¹)	7.97	8.89	11.53	11.01
EC (μs.cm ⁻²)	444	476	497	447
TDS (mg.l ⁻¹)	216	230	239	216
Air T (°C)	31.4	23.8	11.4	9.7
pH	7.75	7.67	7.81	7.75
Salinity	0.21	0.23	0.24	0.22

Table 2. Taxa identified in Golestan waterfall during seasons of 2019 (L: Length, W: Weight, S: Striae, C: Costae, F: Fibula)

Scientific name	Dimensions
Bacillariophyceae	
Achnanthidaceae	
<i>Achnanthidium_grasillimum</i> (F.Meister) Lange-Bertalot	L: 17.5-21 μ m W:3-4 μ m S:14-16
<i>Achnanthidium_minutissimum</i> (Kützing) Czarnecki	L:7.5-13 μ m W:2.5- 3 μ m S:27-30
Amphipleuraceae	
<i>Amphipleura_pellucida</i> (Kützing) Kützing	L:60-93 μ m W:9-11 μ m
<i>Frustulia_vulgaris</i> (Thwaites) De Toni	L:48-49 μ m W: 9-10 μ m
Basillariaceae	
<i>Hantzschia_amphioxys</i> (Ehrenberg) Grunow	L: 35 μ m W:6 μ m F:6
<i>Nitzschia_commutatoides</i> Lange-Bertalot	L: 112 μ m W: 15 μ m F:9-11
<i>Nitzschia_dissipata</i> (Kützing) Rabenhorst	L:20-42 μ m W:3-4 μ m F:7-11
<i>Nitzschia</i> sp.	L: 58-120 μ m W:3-6 μ m F:10-11
<i>Tryblionella_angustata</i> W.Smith	L: 44-52 μ m W: 6-7 μ m S:17-18
<i>Tryblionella_apiculate</i> W.Gregory	L:44-47.5 μ m W:5 μ m S:16-17
<i>Tryblionella_brunoi</i> (Lange-Bertalot) Cantonati & Lange-Bertalot	L:79-83 μ m W:10 μ m S:13
Catenulaceae	
<i>Amphora_inariensis</i> Krammer	L:12-16 μ m W:2.5-4 μ m S:15-17
<i>Amphora_pediculus</i> (Kützing) Grunow	L:10-12.5 μ m W:2.5 μ m S:18
Cocconeidaceae	
<i>Cocconeis_pediculus</i> Ehrenberg	L:27.5 μ m W:19-21 μ m S:16
Cymbellaceae	
<i>Cymbella_affinis</i> Kützing	L:27-28.5 μ m W:7.5-8 μ m S:9-11
<i>Cymbella_compacta</i> Østrup	L: 44 μ m W:14 μ m S:11
<i>Cymbopleura_amphicephala</i> (Nageli) Krammer	L:30-32.5 μ m W:10-11 μ m S:8-10
<i>Cymbopleura_citrus</i> (J.R.carter & Bailey-Watts) Krammer	L: 30 μ m W:10 μ m S: 10
<i>Cymbopleura_kuelbsii</i> Krammer	L:38-32 μ m W:6-7.5 μ m S:11-13
<i>Cymbopleura</i> sp.	L:30-35 μ m W:7-7.5 μ m S:8-10
<i>Cymbopleura</i> cf. <i>vrana</i> Krammer	L: 36 μ m W:10.5 μ m S: 9
Diploneidaceae	
<i>Diploneis_calcilacustris</i> Lange-Bertlot & A.Fuhrmann	L:17-20 μ m W:10-12 μ m S:11-13
<i>Diploneis_krammeri</i> Lange-Beterlot & E.Reichardt	L: 45 μ m W:17-17.5 μ m S: 12
<i>Diploneis_separanda</i> Lange-Beterlot	L:25 μ m W:12 μ m S:12
Fragilariaceae	
<i>Fragilaria_recapitellata</i> Lange-Bertalot & Metzeltin	L:17-18 μ m W:5 μ m S:13-14
Gomphonemataceae	
<i>Delicatophycus_sinensis</i> M.J.Wynne	L:27.5 μ m W:5 μ m S:16
<i>Delicatophycus_verenae</i> M.J.Wynne	L:33-42 μ m W:6-7.5 μ m S:12-14
<i>Encyonopsis_minuta</i> Krammer & E. Reichardt	L:12.5 μ m W:2.5-3 μ m
<i>Gomphonema_micropus</i> Kützing	L: 25 μ m W: 7 μ m S: 11
<i>Gomphonema_parvulum</i> (Kützing) Kützing	L:12-23 μ m W:5-6 μ m S:12-13
<i>Gomphonema_pumilim</i> (Grunow) E.Reichardt & Lange-	L:16-30 μ m W:3-5 μ m S:10-13

Bertalot	
<i>Gomphonema_subclavatum</i> _(Grunow) Grunow	L:37-52 µm W:7-10 µm S:10-12
Naviculaceae	
<i>Gyrosigma</i> _sp.	L:120 µm W:24 µm S:13
<i>Navicula_capitatoradiata</i> _H.Germain ex Gasse	L: 37 W:7.5 S:15-17
<i>Navicula_cryptotenella</i> _Lange-Bertalot	L:16-22 µm W:5-6 µm S:12-14
<i>Navicula_rostellata</i> _Kützing	L: 38 µm W: 9 µm S:12
<i>Navicula_tripunctata</i> _(O.F.Müller) Bory	L:46-52 µm W:8-9 µm S:11-12
Rhoicospheniaceae	
<i>Rhoicosphenai_abbreviata</i> _(C.Agardh) Lange-Bertalot	L:27.5-40 µm W:4-5 µm S:12- 14
Stauroneidaceae	
<i>Craticula_buderi</i> _(Hustedt) Lange-Bertalot	L:20-26 µm W:5-6.5 µm S:18-19
<i>Stauroneis_separanda</i> _Lange-Bertalot & Werum	L: 15-16 µm W: 3.5-4.5 µm
<i>Stauroneis_smithii</i> _Grunow	L:27 µm W:7.5 µm
Surirellaceae	
<i>Surirella_angusta</i> _Kützing	L:23 W:7.5 S:25
<i>Surirella</i> _sp.	L:36 W:24 F: 5-6 C:18
<i>Surirella_librile</i> _(Ehrenberg) Ehrenberg	L:60-88 µm W:20 µm
Tabellariaceae	
<i>Diatoma_moniliformis</i> _(Kützing) D.M.Williams	L:12.5-16 µm W:3.5-4 µm C:7
<i>Halamphora_veneta</i>	L:19-22 µm W:4-5 µm S:16-17
Ulnariaceae	
<i>Ulnaria_ulna</i> _(Nitzsch) Compere	L:102-140 µm W: 6 µm S:10

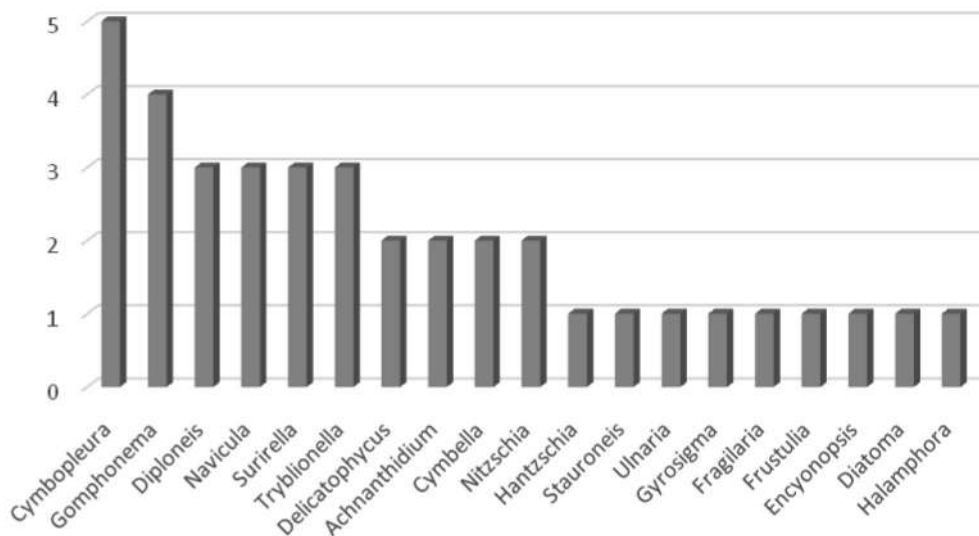


Fig. 2. Number of taxa in each of the genus recognized in the study

Order: Cymbellales

Family: Gomphonemataceae

Genus: *Delicatophycus*

Delicatophycus verenae M.J.Wynne

Synonyms: *Delicata verenae* Lange-Ber-

talot & Krammer

Reference: as *Delicata verenae* in Krammer, 2003, Plate 137, Figs. 1-15.

Description: Valves slightly dorsiventral, elliptic-lanceolate with round not protract-

ed apices, with 33-42.5 μm length and 6-7.5 μm width; axial area widening towards the middle of valve; central area almost absent; raphe lateral, narrowing towards the distal ends; striae radiate, 12-14 /10 μm in middle of valve.

Order: Bacillariales

Family: Bacillariaceae

Genus: *Tryblionella* W. Smith

Tryblionella brunoi (Lange-Bertalot) Canto-nati & Lange-Bertalot

Synonyms: *Nitzschia brunoi* Lange-Bertalot (Powers, 2018; as *Nitzschia brunoi* in Lange-Bertalot and Metzeltin, 1996, Plate 101, Figs. 11-15)

Description: Valves linear with wedge-shaped ends with 78-82.5 μm length and 10-11 μm width; striae are parallel distinctly punctate 13/10 μm .

Order: Naviculales

Family: Stauroneidaceae

Genus: *Stauroneis* Ehrenberg

Stauroneis separanda Lange-Bertalot & Werum

No Synonym

Reference: Bahls, 2012; Levkov et al., 2016, Figs. 48-63.

Description: Valves linear- lanceolate, wider at the center, with triundulate margins, apices rostrate, with the length of 15-16 μm and 3.5-4.5 μm width; central stauros is narrow and linear; pseudoseptum present at the apices; striae fine.

Discussion

In general, most of the previous studies on waterfalls have primarily focused

on tourism, geology, and hydrology, while limited ones have highlighted biodiversity in the ecosystem (Offem and Ikpi, 2012). Therefore, waterfalls are sometimes considered as lifeless zones (Chernicoff et al., 1997). The present study aimed to identify epilithic diatoms in Golestan waterfall.

Reviewing the studies conducted on diatom in Golestan province demonstrated 80% similarity between the results of the present study with those of Ahmadi et al. (2019) regarding the diatoms of three springs. In addition, the taxa of *Amphipleura pellucida*, *Diploneis calcilacustris*, *D. krammeri*, *D. separanda*, *Fragilaria recapitellata*, *Frustulia vulgaris*, *Gomphonema subclavatum*, *Stauroneis smithii*, and *Surirella angusta* were found only in these two studies. However, the others except for new records were reported in the previous research.

Among the most abundant taxa, *A. minutissimum*, and *C. affinis* were abundant in most ecosystems of the province (Aghatabay, 2018; Bayani, 2019; Dadgar, 2016; Lakzaie, 2016). Further, *A. minutissimum* is considered as one of the most frequently recorded taxa worldwide (Falasco et al., 2012, 2016; Kelly et al., 2007; Kheiri et al., 2019; Krammer and Lange-Bertalot, 1991b;) from oligo- to hypertrophic in alkaline to acidic waters (Potapova and Hamilton, 2004). Although Van Dam et al. (1994) classified *C. affinis* as eutrophic taxon, BCG classification indicated human disturbance (Davies and Jackson,

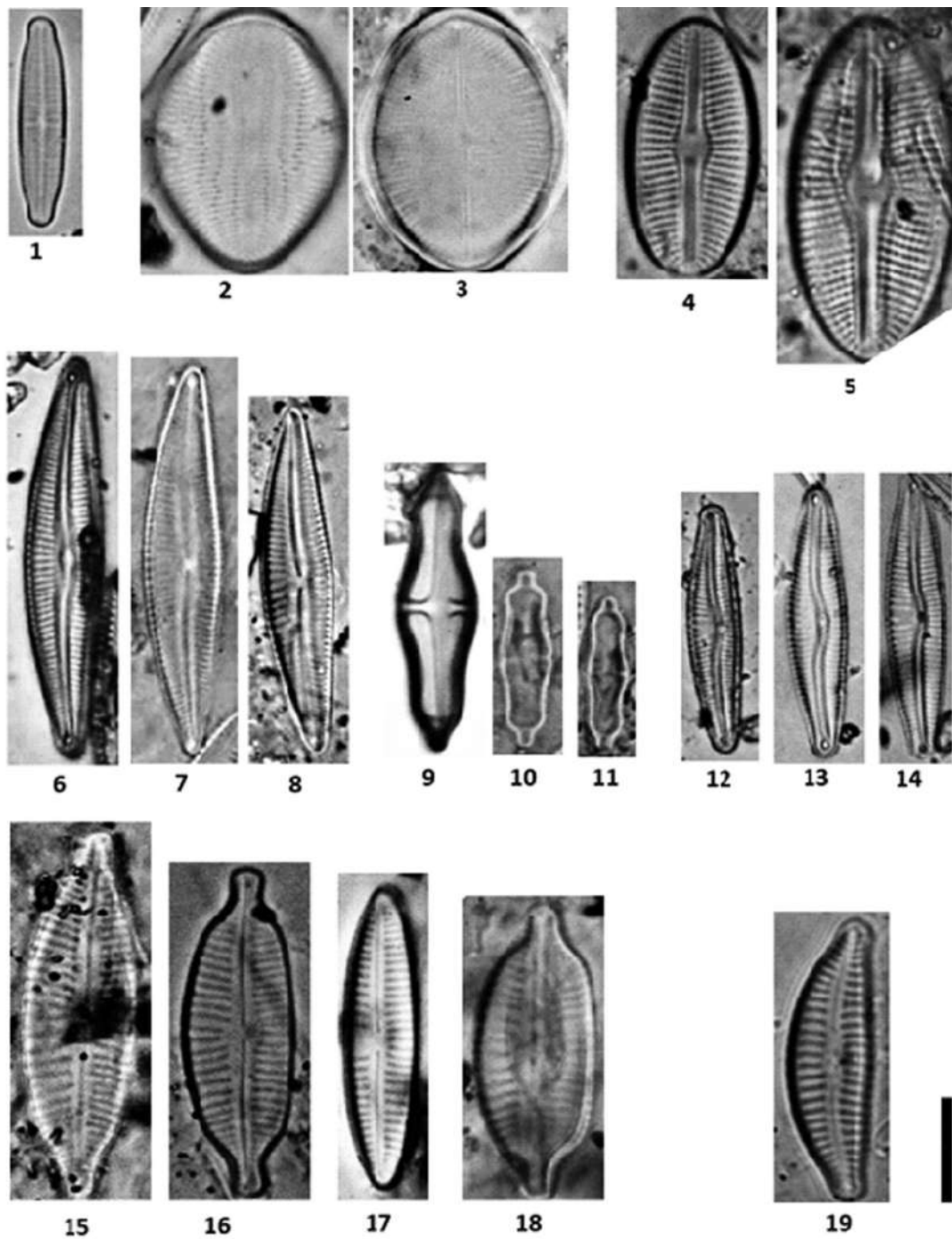


Plate I. (1-20). (1) *Achnantheidium gracillimum*, (2,3) *Cocconeis pediculus*, (4) *Diploneis separanda*, (5) *Diploneis krammeri*, (6-8) *Delicatophycus verenaë*, (9) *Stauroneis smithii*, (10, 11) *Stauroneis separanda*, (12-14) *Delicatophycus sinensis*, (15) *Cymbopleura* cf. *irana*, (16) *Cymbopleura amphicephala*, (17) *Cymbopleura kuelbsi*, (18) *Cymbopleura citrus*, (19) *Cymbella affinis*. Bar: 10 μ m.

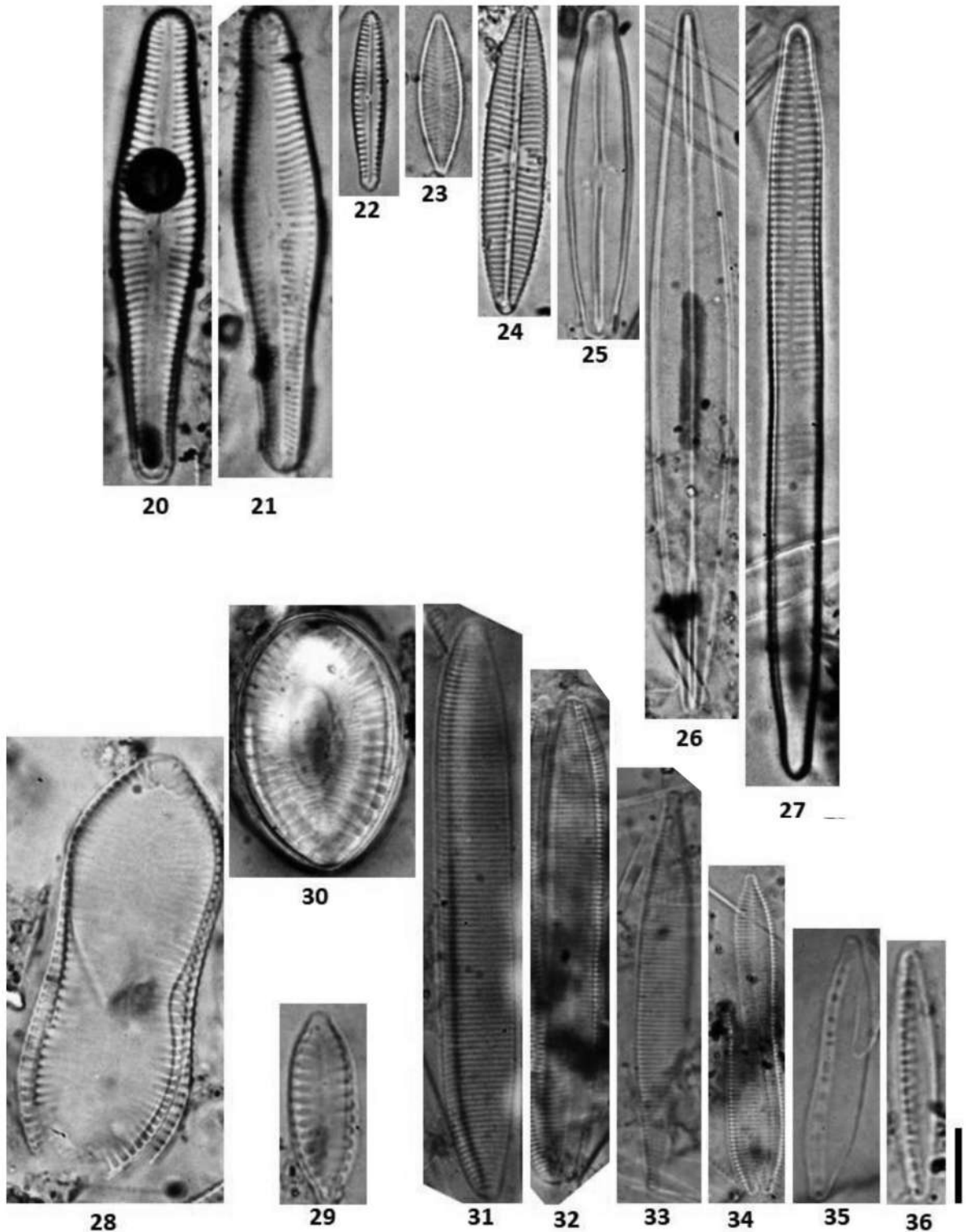


Plate II. (20-36). (20, 21) *Gomphonema subclavatum*, (22) *Gomphonema pumilum*, (23) *Navicula cryptotenella*, (24): *Navicula tripunctata*, (25) *Frustulia vulgaris*, (26). *Amphipleura pellucida* (27) *Ulnaria uln*, (28) *Surirella libril*, (29) *Surirella angusta*, (30) *Surirella* sp., (31,32) *Tryblionella brunoi* (33) *Tryblionella angustata*, (34) *Tryblionella apiculata*, (35) *Hantzschia amphyoxys* (36) *Nitzschia dissipata*. Bar: 10 μ m.

2006) characterized *C. affinis* as highly sensitive species (Potapova, 2011) in California State. *C. affinis*, a common taxon both in rivers and lakes (Krammer and Lange-Bertalot, 1986; Patrick and Reimer, 1975), is abundant in oligotrophic and mineralized systems (Tornes, et al., 2007) and neighbor countries (Khazal et al., 2018). Further, *A. pellucida* is an oligo-mesotrophic species (Van Dam et al., 1994), as well as a cosmopolitan alkaliphilous taxon, which is found in pH 6.2-8 (Lowe, 1974). Additionally, the species can be observed as planktonic despite its existence as benthic taxon in most cases (Krizmanic et al. 2008). Although *A. pellucida* is a widely-distributed taxon, it was only reported from Kashkan River (Safiallah et al., 2020) and Gole-Ramian spring with low abundances in Iran (Ahmadi et al., 2019). Furthermore, *Cymbopleura kuelbsii* firstly recorded by Kheiri et al (2019), this taxon is newly initiated species (Krammer, 2003) and there is not more data on its ecology, however in our study *C. kuelbsii* was abundant only in winter. The water was well oxygenated with low nutrient and moderate conductivity.

Assymetric biraphid was determined as the most diverse group, which is in line with the results of the previous research in Golestan province (Aghatabay, 2018; Bayani, 2019; Dadgar, 2016; Lakzaie, 2016), while they are inconsistent with those related to

Kashkan River in Zagros (Safiallah et al., 2020). In the present study, the genera of the group included *Amphora*, *Cymbella*, *Cymbopleura*, *Delicatophycus*, *Encyonopsis*, *Gomphonema*, *Rhoicosphenia* and *Halamphora*. Further, symmetrical biraphid (*Amphipleura*, *Craticula*, *Diploneis*, *Frustulia*, *Stauroneis*, and *Navicula*) Araphids (*Diatoma*, *Fragilaria*, and *Ulnaria*), Nitzschioid (*Hantzschia*, *Nitzschia*, and *Tryblionella*), monoraphids (*Achnanthisidium* and *Cocconeis*), and Surirelloid (*Surirella*) were present in the waterfall. Unlike the previous studies, centric group had no representative in our study along with Epithemioid and Eunotioid groups.

Among the diatom genera, *Nitzschia*, *Navicula*, and *Gomphonema*, as large genera with many species, have been usually reported as the most species-rich genera in the studies conducted in Iran (Ahmadi et al., 2019; Mehrani Adl, 2020; Panahy et al., 2018) and the world (Jakivljevic et al., 2016; Noga et al., 2013; Solak, 2011). Furthermore, *Cymbopleura* was obtained as the most species-rich genus, which is not in line with the results of other studies in Golestan province, and above-mentioned ones. Among the species of this genus identified in our study *C. citrus* for the first time introduced by Aghatabay (2018) and Ahmadi et al (2019) to diatom flora of Iran from Golestan Province. Other species of *Cymbopleura* in-

cluding *C. cf. vrana* have not been reported from Golestan Province yet, even though were recorded from Karaj River by Kheiri et al (2019). *Gomphonema* and *Navicula* were among the major genera too, however unlike their studies, *Nitzschia* was less important in our work.

In this research, three taxa were recorded for the first time for diatom flora of Iran. *Delicatophycus* established as a new genus, for substituting genus *Delicata* which was declared invalid by Wynne (2019). Indeed, *Delicatophycus sinensis* that were recorded by Kheiri et al (2019) as *Delicata sinensis* from Karaj River as new record for Iran, and *Delicatophycus* in which there is no record in diatom flora of Iran. Additionally, *D. verena* was reported as a new species only by Krammer (2003) and no data are available on the ecology of the species. The results of the present study demonstrated the above-mentioned species of the genus in alkaline water with low conductivity, and nutrients, both of which were abundant in samples, especially *D. Sinensis*. *D. verena* was introduced for the first time.

Further, genus *Tryblionella* was determined as another new record, which was described by Smith (1853), and many of its species were derived from the large genus *Nitzschia* by Round et al. (1990). Regarding *Tryblionella*, raphe is eccentric, similar to that of *Nitzschia*, while *Tryblionella* has longitudi-

nal undulations on valve face. Furthermore, *Tryblionella brunoi* (T. brunoi) was reported in meso-to oligotrophic waters, as well as eutrophic ones in a few cases (Powers, 2018).

Stauroneis is a genus from symmetrical biraphid group with naviculoid cells. Further, the main characteristics of the genus are presence of stauros in the valve center and punctate striae. Additionally, Most species of this genus are characteristic of oligotrophic waters in the temperate zone (Lange-Bertalot and Metzeltin, 1996). However *S. separanda* is distinguished by smaller valves and low undulation in outline, is similar to *S. smithii*. Furthermore, this taxon was recorded firstly by Werum and Lange-Bertalot (2004), as widely distributed in calcareous springs with high conductivity in Europe. While Levkov et al. (2016) recorded this species from mesotrophic calcareous waters from Macedonia. Moreover, this taxon was observed in co-occurrence with *S. smithii* that is in accordance with present study.

Given that Iran is considered as a vast country with diverse aquatic systems, such local and detailed studies can result in improving the knowledge on diatom flora in Iran. In fact, the present study is the first research on the diatom diversity of waterfall in Iran. In general, the diatom flora of Golestan Waterfall included some cosmopolitan taxa found in various habitats, along with taxa with

narrow distribution. The presence of taxa with limited distribution worldwide indicated the importance of conserving waterfalls. Accordingly, further studies should be conducted in such ecosystems. Due to the tourism value of waterfalls, they are subjected to human impacts and environmental disturbances. Thus, the protection of waterfall habitats is important for conserving aquatic diversity.

Acknowledgment

Authors would like to thank Gonbad Kavous University for financial support.

References

- Aghatabay A. (2018). Investigation of the epilithic and epipelagic diatoms in the Khormarud River of Azadshahr, Golestan Province. MSc thesis, GonbadKavous University, GonbadKavous, Iran (in Persian).
- Ahmadi Musaabad L, Panahy Mirzahasanlou J, Mahmoodlu MG, Bahlakeh A. (2019). Diatom flora in three springs of Golestan Province. *Journal of Phycological Research*. 3 (2): 432-442.
- Andrejic JC, Krizmanic J, Cvijan M. (2012). Diatom species composition of the Nisava River and its tributaries Jerma and Temska Rivers (southern Serbia). *Archive of Biological Sciences*. 64 (3): 1127-1140. Doi: 10.2298/ABS1203127A.
- Bahls L. (2006). Northwest Diatoms. A photographic catalogue of species in the Montana diatom collection. Montana diatom collection.
- Bahls L. (2012). *Stauroneis separanda*. In diatoms of North America. https://diatoms.org/species/stauroneis_separanda.
- Bahls L. (2019). Diatoms from Western North America. 2. *Cymbella falsa*, *Cymboplectura* and *Delicatophycus* (Bacillariophyta). Taxonomy, Ecology, Biogeography. The Montana diatom collection. 114 pp.
- Bayani M. (2019). Investigation of the Epilithic Diatoms in GharahChay River of Ramian in Golestan province. MSc thesis, Gonbad Kavous University, Gonbad Kavous, Iran (in Persian).
- Bellinger EG and Sigeo DC. (2010). Freshwater algae, identification and use as bioindicators. First ed. Wiley-Blackwell Press, UK.
- Chernicoff S, Fox H, Venkatakrishnan R. (1997). *Essentials of Geology*. Worth Publishers, New York.
- Clesceri L, Greenberg AE, Eaton AD. (1999). Standard methods for examination of water and wastewater. American public health association, American water works Association, Water environment federation. Waldorf, Maryland.
- Dadgar A. (2016). Investigation of the epilithic diatoms in the Zarringol River of Golestan Province. MSc thesis, Gonbad Kavous University, Gonbad Kavous, Iran (in Persian).
- Davies SP and Jackson SK. (2006). The biological condition gradient: a descriptive model for interpreting change in aquatic ecosystems. *Ecological Applications*. 16 (4): 1251-66. Doi: 10.1890/1051-0761(2006)016[1251:tb-cgad]2.0.co;2
- Delgado C, Pardo I, Gracia L. (2012). *A multi-metric diatom index to assess the ecological*

- status of coastal Galician rivers (NW Spain). *Hydrobiologia*. 644: 371-384. Doi: 10.1007/s10750-010-0206-y.
- Falasco E, Ector L, Ciaccio E, Hoffmann L, Bona F. (2012). Alpine freshwater ecosystems in a protected area: a source of diatom diversity. *Hydrobiologia*. 695: 233-251. Doi: 10.1007/s10750-012-1114-0.
- Falasco E, Piano E, Bona F. (2016). Diatom flora in Mediterranean streams: flow intermittency threatens endangered species. *Biodiversity and Conservation*. 25: 2965-2986. Doi: 10.1007/s10531-016-1213-8.
- Jakovljevic OS, Popovic SS, Vidakovic DP, Stojanovic KZ, Krizmanic JZ. (2016). The application of benthic diatoms in water quality assessment (Mlav River, Serbia). *Acta Botanica Croatica*. 75 (2): 199-205. Doi: 10.1515/botcro-2016-0032.
- Krizmanic J, Rzanicanin A, Subakov-Simic G, Cvijan M. (2008). *Amphipleura pelucida* (Kutz.) Kutz.- An emended diagnosis concerning valve length. *Diatom Research*. 23 (1): 243-248. Doi: 10.1080/0269249X.2008.9705750.
- Khazal D, Al-Jibouri W, Hassan FM, Hakman AA. (2018). Ecological and taxonomical study of epipelon community in Diyala River in Diyala Province-Iraq. *Indian Journal of Ecology*. 45(4): 680-688.
- Kelly M G, Juggins S, Bennion H, Burgess A, Yallop M, Hirst H, King L, Jamieson BJ, Guthrie R, Rippey B. (2007). Use of diatoms for evaluating ecological status in UK freshwaters. Science Report: SC030103. Environment Agency, Bristol.
- Kheiri S, Solak CN, Edlund MB, Spaulding S, Nejadstari T, Asri Y, Hamdi SMM. (2019). Biodiversity of diatoms in the Karaj River in the Central Alborz, Iran. *Diatom Research*. 33 (3): 355-380. Doi: 10.1080/0269249X.2018.1557747.
- Krammer K. (2002). *Cymbella*. Diatoms of Europe. Diatoms of the European inland waters and comparable habitats. 3: 1-584.
- Krammer K. (2003). *Cymbopleura, Delicata, Navicymbula, Gomphocymbellopsis, Afro-cymbella*. Diatoms of Europe 4: 1-530.
- Krammer K and Lange-Bertalot H. (1986). Bacillariophyceae, 1. Naviculaceae. In: Etti, H. Gerloff, J. Heyning, H. Mollenhauer, D. (eds), *Susswasserflora von Mitteleuropa*. Vol.1. Gustav Fischer Verlag. Jena.
- Krammer K and Lange-Bertalot H. (1988). Bacillariophyceae, 2. Bacillariaceae, Epithemiaceae, Surirellaceae. In: Etti, H. Gerloff, J. Heyning, H. Mollenhauer, D. (eds), *Susswasserflora von Mitteleuropa*. Vol. 2. Gustav Fischer Verlag. Stuttgart.
- Krammer K and Lange-Bertalot H. (1991a). Bacillariophyceae, 3. Centrales, Fragilariaceae, Eunotiaceae. In: Etti, H. Gerloff, J. Heyning, H. Mollenhauer, D. (eds), *Susswasserflora von Mitteleuropa*. Vol. 3. Gustav Fischer Verlag. Stuttgart.
- Krammer K and Lange-Bertalot H. (1991b). Bacillariophyceae, Achnanthaceae. Kritische Ergänzungen zu *Navicula* (Lineolatae) und *Gomphonema*. In: Etti, H. Gerloff, J. Heyning, H. Mollenhauer, D. (eds), *Susswasserflora von Mitteleuropa*. Vol. 4. Gustav Fischer Verlag. Stuttgart.
- Lakzaie F, Panahy Mirzahasanlou J, Gholizadeh M, Daneshvar A. (2018). Ecological

- study of the diatoms in ChehelChay River of Minudasht in Golestan province. *Journal of Aquatic Production and Utilization*. 7 (3): 41-51. (In Persian).
- Lange-Bertalot H and Metzeltin D. (1996). Indicators of oligotrophy. 800 taxa representative of three ecologically distinct lake types, carbonate buffered-Oligodystrophic-weakly buffered soft water with 2428 figures on 125 plates. Koeltz Scientific Books, Germany.
- Levkov Z, Tofilovska S, Jovanovska E, Cvetkoska A, Metzeltin D. (2016). Revisión of the *Stauroneis smithii* Grunow (Bacillariophyceae) species complex from Macedonia. *Botanica Serbica*. 40(2): 167-178.
- Lowe RL. (1974). Environmental requirements and pollution tolerance of freshwater diatoms. United States Environmental Protection Agency, Cincinnati, OH. 333 pp.
- Martin G, Toja J, Sala SE, Fernandez MR, Reyes I, Casco MA. (2010). Application of diatom biotic indices in the Guadalquivir River basin, a Mediterranean basin. Which one is the most appropriated? *Environmental Monitoring Assessment*. 170: 519-534. Doi: 10.1007/s10661-009-1254-5.
- MehraniAdl M, Iranbakhsh A, Noroozi M, Asri Y, Saadatmand S. (2020). Epipellic diatoms flora of Kordan River, Alborz province in Iran. *Modern Phytomorphology*. 14: 40-48.
- Noga T, Stanek-Tarkowska J, Kochman N, Peszek L, Pajaczek A, Wozniak K. (2013). Application of diatoms to assess the quality of the waters of the Baryczka Stream, left-side tributary of the River San. *Journal of Ecological Engineering*. 14: 8-23. Doi: 10.5604/2081139X.1055818.
- Offem BO and Ikpi GU. (2012). Distribution and dynamics of a tropical waterfalls ecosystems. *International Journal of Ecosystem*. 2 (1): 28-37. Doi: 10.5923/j.ije.20120201.05.
- Panahy Mirzahaslou J, Nejadstarrari T, Ramezanpour Z, ImanpourNamin J, Asri Y. (2018). The epilithic and epipellic diatom flora of the Balikhli River, Northwest Iran. *Turkish Journal of Botany*. 42: 518-532. Doi: 10.3906/bot-1711-46.
- Patrick RR and Reimer CW (1975). The diatoms of the United States. Vol. 1. The Academy of Natural Sciences of Philadelphia.
- Potapova M. (2011). *Cymbella affinis*. In diatoms of North America. Retrieved January 2, 2021 <https://diatoms.org/species/cymbella-affinis>.
- Potapova M and Hamilton PB. (2007). Morphological and Ecological variation within the *Achnantheidium minutissimum* (Bacillariophyceae) species complex. *Journal of Phycology*. 43: 561-575. Doi: 10.1111/j.1529-8817.2007.00332.x.
- Powers M. (2018). *Tryblionella brunoi*. In diatoms of North America. Retrieved January 2, 2021. <https://diatoms.org/species/tryblionella-brunoi>.
- Rawat US and Agarwal NK. (2015). Biodiversity: Concept, threats and conservation. *Environment Conservation Journal*. 16 (3): 19-28.
- Round FE, Crawford RM, Mann DG. (1990). The diatoms. Biology and morphology of the genera. Cambridge University Press, Cambridge, UK. 747 pp.
- Safiallah S, Saadatmand S, Kheiri S, Iranbakhsh A. (2020). Biodiversity of diatoms in the Kashkan River in the Zagros Moun-

- tains, western Iran. Iranian Journal of Botany. 206 (2): 141-161. Doi: 10.22092/ijb.2020.351698.1296.
- Smith W. (1853). A synopsis of the British Diatomaceae. Vol. 1. John van Voorst, London.
- Solak NV. (2011). The application of diatom indices in the upper Porsuk Creek Küta-hya-Turkey. Turkish Journal of Fisheries and Aquatic Sciences. 11: 329-337. Doi: 10.4194/trjfas.2011.0105.
- Taylor JC, Harding WR, Archibald CGM. (2007). A methods manual for the collection, preparation and analysis of diatom samples. ver. 1. 49 pp. Water Research Commission.
- Tornes E, Cambra J, Goma J, Leira M, Ortiz R, Sabater S. (2007). Indicator taxa of benthic diatom communities: a case study in Mediterranean streams. International Journal of Limnology. 43 (1): 1-11. Doi: 10.1051/limn/2007023.
- Van Dam H, Mertens A, Sinkeldam J (1994). A coded checklist and ecological indicator values of freshwater diatoms from the Netherlands. Netherlands Journal of Aquatic Ecology. 26: 117-133. Doi: 10.1007/BF02334251.
- Werum M and Lange-Bertalot H. (2004). Diatoms in springs from central Europe and elsewhere under the influence of hydrogeology and anthropogenic impacts. Iconographia Diatomologica 13:1-417.
- Wynne MJ. (2019). *Delicatophycus* gen. nov.: a validation of “*Delicata* Krammer” inval. (Gomphonemataceae, Bacillariophyta). Notulae Algarum. 97: 1-3.